

# Ser-249 p53 Mutations in Plasma DNA of Patients With Hepatocellular Carcinoma From The Gambia

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**Background:** A selective mutation, an arginine-to-serine substitution in codon 249, of the p53 gene has been identified as a "hotspot" mutation in hepatocellular carcinoma (HCC). This mutation occurs in populations that are exposed to aflatoxins and have a high prevalence of hepatitis B virus carriers. We evaluated whether this mutation could be detected in cell-free DNA isolated from the plasma of subjects from The Gambia to detect this mutation that is strongly associated with HCC. **Methods:** Fifty-three patients with HCC, 13 patients with cirrhosis, and 53 control subjects were prospectively recruited from The Gambia. Sixty patients, of non-African origin, with various liver pathologies were also selected from France. DNA was extracted and purified from 200- $\mu$ L aliquots of plasma. The Ser-249 p53 mutation was detected by restriction endonuclease digestion of polymerase chain reaction products from exon 7 and was confirmed by direct sequencing of the amplified DNA. **Results:** The Ser-249 p53 mutation was detected in plasma DNA from 19 (36%) of the 53 patients with HCC, two (15%) of the 13 patients with cirrhosis, and three (6%) of the 53 control subjects. This mutation was not detected in any plasma DNA from the European patients. The adjusted odds ratio for having the mutation was 16.4 (95% confidence interval = 3.0–90.5) for patients with HCC compared with the control subjects. **Conclusion:** The Ser-249 p53 mutation in plasma DNA is strongly associated with HCC in Gambian patients. This mutation was also detected at a much lower prevalence in plasma DNA from Gambian patients with cirrhosis and in Gambian control subjects, findings that may lead to the

earlier detection of HCC. Use of the Ser-249 p53 mutation should facilitate further molecular epidemiologic studies on the development of HCC. [J Natl Cancer Inst 2000;92:148–53]

In sub-Saharan Africa and Asia, hepatocellular carcinoma (HCC) is a major cause of cancer death (1). The primary etiologic agents identified in these regions are chronic infection with hepatitis B virus (HBV) and dietary exposure to aflatoxin B<sub>1</sub> (AFB<sub>1</sub>). Epidemiologic evidence and experimental evidence support the association of HBV (2) and AFB<sub>1</sub> (3) with an increased risk of HCC and suggest that combined exposure may act synergistically (4–6).

Tumor-specific p53 mutations have been identified in several human cancers (7,8). A selective guanine-to-thymine transversion mutation in codon 249 (AGG to AGT [transversion underlined] leading to an arginine-to-serine substitution) of the p53 gene (also known as TP53) has been identified as a "hotspot" mutation for HCC (9,10). Epidemiologic evidence and experimental evidence have suggested that in HCC this mutation is strongly associated with exposure to AFB<sub>1</sub> (6,7,11).

The Gambia in West Africa has a population with a high incidence of HCC, a high level of endemic, chronic HBV infection, and a high exposure to AFB<sub>1</sub>. Data from the Gambian National Cancer Registry indicate that HCC is the most common cancer in males and the second most common cancer in females (12,13). Ten percent to 20% of the Gambian population is chronically infected with HBV (14,15). Extensive analysis of individual biomarkers of aflatoxin exposure in The Gambia has demonstrated ubiquitous dietary exposure, with more than 95% of the population having detectable levels of aflatoxin-albumin adducts in their serum (6,11).

Recent research has shown that DNA can be isolated from the plasma or serum of patients with cancer; this plasma DNA carries the same genetic mutations as DNA in the tumor. Thus, plasma DNA can be used as surrogate material to detect genetic alterations present in the original tumor (16–18). We have used this method to determine whether Ser-249 p53 mutations can be detected in plasma DNA isolated from Gambian patients with HCC, Gambian patients with cirrhosis, and healthy Gambian control subjects, as well

as from European patients with HCC or with cirrhosis.

## MATERIALS AND METHODS

### Subjects and Specimens

Incident cases of HCC and cirrhosis were identified from liver disease referral clinics at each of the three tertiary referral hospitals in The Gambia (i.e., Royal Victoria Hospital, Medical Research Council Hospital, and Bansang Hospital). Samples from The Gambia analyzed in this study were from subjects sequentially recruited from September 1997 through June 1998 into an ongoing case-control study of HCC. We amplified DNA from 142 (92%) of 155 subjects enrolled during this period; we could not amplify DNA from 13 subjects. Evaluation of subjects included a clinical examination, ultrasonography, collection of biologic specimens, and a structured interview.

For inclusion in the study, a patient with HCC had to have compatible clinical and ultrasonographic findings and serum  $\alpha$ -fetoprotein levels of 100 ng/mL or more. Twenty-three patients with HCC, who were initially recruited into the study, were excluded because their levels of  $\alpha$ -fetoprotein were less than 100 ng/mL. Analyses performed with and without inclusion of these subjects were similar. The results presented include only patients who met the case definition of HCC, for a total of 119 study subjects (53 patients with HCC, 13 patients with cirrhosis, and 53 control subjects). Patients with cirrhosis were included as an additional referent group, in addition to healthy control subjects, to evaluate factors associated with the progression to HCC. The diagnosis of cirrhosis was based on compatible clinical history and ultrasonographic patterns characteristic of cirrhosis, without any focal lesions suggestive of HCC. A minority of patients with liver disease also had histologic confirmation. Control subjects were individuals with no history or clinical findings suggestive of liver disease, who were recruited from the outpatient clinics of one of the three tertiary referral hospitals. Control subjects were frequency matched to case patients with HCC by age (within 10-year groupings), sex, and recruitment site. Local and international ethical and scientific review committees approved the study protocol, and informed consent

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was obtained from each participant before inclusion in the study.

To compare West African subjects with subjects from areas with lower exposure to aflatoxin and HBV, we evaluated plasma and serum samples obtained from 60 non-African patients from Europe (Hotel Dieu Hospital [Lyon, France] and Necker Hospital [Paris, France]). Subjects included 50 patients with HCC (21 associated with HBV, 17 with hepatitis C virus, seven with alcohol, and five with unknown status) and 10 patients with cirrhosis (three associated with HBV, four with hepatitis C virus, and seven with alcohol [some patients with more than one association]). The samples were sent to the laboratory of the International Agency for Research on Cancer (IARC) and were analyzed in the same manner as the samples from The Gambia.

## DNA Extraction and Polymerase Chain Reaction

Blood specimens anticoagulated with EDTA were processed immediately after collection and stored at  $-70^{\circ}\text{C}$  at the Medical Research Council Laboratories, The Gambia, until subsequent testing.  $\alpha$ -Feto-protein and HBV serologic testing was performed with standard laboratory kits. A 500- $\mu\text{L}$  aliquot of plasma was shipped on dry ice to IARC laboratories (Lyon, France) to test for the Ser-249 p53 mutation. DNA was extracted from 200  $\mu\text{L}$  of plasma with a QiAmp tissue kit (Qiagen, Hilden, Germany). The purified DNA was eluted from the QuiAmp silica column with one volume (50  $\mu\text{L}$ ) of nuclease-free water (Promega Corp., Madison, WI).

Ten microliters of DNA eluate was used to amplify exon 7 of the p53 gene with primers located in the introns flanking exon 7, as described by Lehman et al. (19). The following primers (Genset, Paris, France) were used: p1 (sense), 5'-CTTGCCACAG-GTCTCCCAA-3'; p2 (antisense), 5'-AGGGGT-CAGCGCAAGCAGA-3'.

The 50- $\mu\text{L}$  reaction mixture contained 50 mM KCl, 20 mM Tris-HCl (pH 8.4), 1.5 mM  $\text{MgCl}_2$ , all four deoxynucleoside triphosphates (each at 0.2 mM; Promega Corp.), 0.2  $\mu\text{M}$  of each primer, and 2.5 U of Platinum *Taq* DNA polymerase (Life Technologies, Inc. [GIBCO BRL], Gaithersburg, MD). After an incubation ( $94^{\circ}\text{C}$  for 2 minutes), 35 cycles ( $94^{\circ}\text{C}$  for 30 seconds,  $60^{\circ}\text{C}$  for 30 seconds, and  $72^{\circ}\text{C}$  for 30 seconds) were performed and were followed by a final 5-minute extension at  $72^{\circ}\text{C}$  in a thermal cycler (gene Amp PCR system, 9600; The Perkin-Elmer Corp., Foster City, CA). The amplification products (254 base pairs [bp]) were visualized by staining with ethidium bromide after electrophoresis on 3% agarose gel.

In six samples, the polymerase chain reaction (PCR) products were not clearly detected, and a second amplification reaction was performed with the following nested primers: p3 (sense), 5'-AGGC-GCACTGGCCTCCTT-3'; p4 (antisense), 5'-TGTGCAGGGTGGCAAGTGGC-3'. This reaction mixture contained 5  $\mu\text{L}$  of the first PCR products, and 25 cycles (as in the first PCR) were performed. The same nested PCR protocol was also used to generate enough material for DNA sequence analysis (see below). All sets of PCRs included negative control samples (containing no template DNA). For avoidance of PCR contaminant artifacts, routine precautions included performing DNA extraction, PCR

assembly, and the PCR in separate rooms and using barrier tips at all stages of the procedure.

## Mutation Detection by Restriction Endonuclease Analysis

PCR products (10  $\mu\text{L}$ ) were digested with restriction endonuclease *Hae*III (Boehringer Mannheim GmbH, Mannheim, Germany), according to the manufacturer's instructions. This enzyme cleaves a GG/CC sequence between codon 249 and codon 250 to generate two fragments of 92 bp and 66 bp (plus several small fragments) from the 254-bp product of the first PCR. The presence of a mutation in codon 249 or codon 250 results in an uncleaved 158-bp fragment. These fragments were separated and identified on 3% agarose gel stained with ethidium bromide (Fig. 1). The absence of the 254-bp band (full-length PCR products) provides a control for complete digestion of the PCR product. Restriction endonuclease assays included both positive (DNA with a Ser-249 p53 mutation isolated from a patient with liver cancer) and negative (wild-type DNA) controls. All reactions were carried out in duplicate. As a further control, five positive and five negative assays were selected at random and submitted to a second, independent analysis, including DNA extracts, PCR, and restriction endonuclease digestion. Concordant results were obtained in each case.

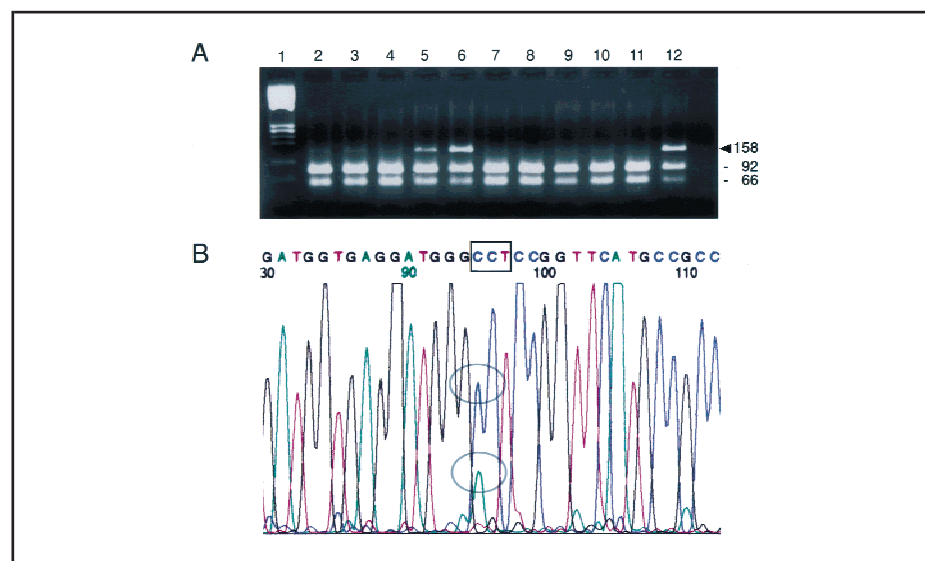
For the determination of the relative sensitivity of this method to detect a mutation, a control DNA sample containing a mutation at codon 249 was serially diluted 1:2 up to 1:256 in wild-type DNA. The mixture was amplified by PCR as described above and analyzed by restriction endonuclease di-

gestion. The presence of the uncleaved 158-bp fragment was clearly visible at ratios of mutant to wild-type DNA of 1:64.

The 158-bp product, not cleaved between codon 249 and codon 250, was excised from agarose gels and further amplified with nested PCR primers P3 and P4, as described above. The products of this second PCR were purified with QIA quick-spin columns (Qiagen), and 0.25  $\mu\text{g}$  of DNA (exon 7, codons 225-260) was analyzed by automated DNA sequencing (sequencer 377; The Perkin-Elmer Corp.) with the use of the dRhodamine terminator cycle sequencing kit (Applied Biosystems, Foster City, CA) (Fig. 1).

## Statistical Analysis

Frequency tables of independent variables and Ser-249 p53 mutations, as a dichotomous variable, were evaluated for statistical significance by Pearson's  $\chi^2$  test and Fisher's exact test. For the estimation of the risk of mutations among the different groups in the study while adjustment was made for potential confounders (including age, sex, recruitment site, and hepatitis B surface antigen [HBsAg] status, a marker of chronic infection with HBV), a multivariable logistic regression analysis was performed to estimate odds ratios (ORs) and 95% confidence intervals (CIs). In a separate stratified analysis, only data from patients with HCC were evaluated by similar methods to examine factors associated with Ser-249 p53 mutation-positive HCC compared with Ser-249 p53 mutation-negative HCC. All analyses were performed with STATA version 5.0 (Stata Corporation, College Station, TX) and EPI-Info version 6.0 (Centers for Disease Control and Prevention, Atlanta, GA; and World Health



**Fig. 1.** Example of detection of plasma Ser-249 p53 mutation in patients with hepatocellular carcinoma (HCC) and control subjects from The Gambia. **A)** p53 exon 7 was amplified by polymerase chain reaction, and mutations at codon 249 were identified by restriction endonuclease digestion (codon 249 contains a *Hae*III site that is destroyed by mutation). The presence of an undigested, 158-base-pair (bp) fragment (arrowhead) is indicative of mutation. The 92-bp and 66-bp fragments are from the wild-type sequence. **Lane 1** = molecular weight markers; **lanes 2-7** = patients with HCC; **lanes 8-11** = control subjects; and **lane 12** = positive control. **B)** Automated DNA sequencing of p53 exon 7 corresponding to the HCC material analyzed above in lane 6. The sequence of codon 249 is enclosed in the box. Peaks corresponding to wild-type (C) or mutant (A) sequence at the first base of codon 249 are circled. Only a portion of the electropherogram is shown.

## RESULTS

The characteristics of the 179 subjects evaluated from The Gambia and Europe are shown in Table 1. The age and sex distributions of Gambian patients with HCC and Gambian control subjects were similar. Among Gambian subjects, patients with cirrhosis were markedly younger than patients with HCC but had a similar sex ratio. Sixty-four percent of Gambian patients with HCC, 54% of Gambian patients with cirrhosis, and 17% of Gambian control subjects were chronic carriers of HBV ( $P < .001$  for difference between groups). As expected, the age distribution of European patients with HCC or with cirrhosis was considerably older, and the prevalence of HBV carriers was lower than that in The Gambia. The age difference between European patients with cirrhosis and European patients with HCC and the sex ratios observed among European subjects were similar, although less pronounced, compared with those observed among Gambian subjects.

A Ser-249 p53 mutation was present in 19 of 53 patients with HCC, two of 13 patients with cirrhosis, and three of 53 control subjects (Table 2), giving a prevalence of 36%, 15%, and 6%, respectively ( $P < .001$  for difference between groups). In univariate analysis stratified by subject

group, there was no statistically significant difference in mutation prevalence by age, recruitment site, or HBsAg status. However, females were statistically significantly more likely to have the Ser-249 p53 mutation than males among both patients with HCC ( $P = .012$ ) and control subjects ( $P = .029$ ). No Ser-249 p53 mutation was observed in 60 sera/plasma samples from European patients with either HCC or cirrhosis.

In multivariable analysis adjusting for age, sex, recruitment site, and HBsAg status, patients with HCC were 16-fold more likely to carry the Ser-249 p53 mutation (OR = 16.4; 95% CI = 3.0–90.5) compared with control subjects (Table 2). Patients with cirrhosis also had an increased risk compared with control subjects (OR = 3.2) but with a wide 95% CI that included unity (95% CI = 0.4–26.1). In this model, female gender was statistically significantly associated with Ser-249 p53 mutations, with an adjusted OR of 4.6 (95% CI = 1.3–16.4) compared with males.

When the analysis among only patients with HCC was stratified by mutation status, no difference in Ser-249 p53 mutation prevalence was observed by age or HBsAg status (Table 3). Females were again overrepresented in the Ser-249 p53 mutation-positive group; however, after adjustment for age, recruitment site, and HBsAg status, the strength of the associa-

tion was of borderline statistical significance ( $P = .053$ ).

## DISCUSSION

We report the presence of Ser-249 p53 mutations in DNA isolated from the plasma of individuals from a West African population at high risk for HCC, exposure to the carcinogen AFB<sub>1</sub>, and chronic HBV infection. This “hotspot” mutation was detected in 36% of patients with HCC (Table 2), with a much lower prevalence in patients with cirrhosis and control subjects. No such mutation was present in the plasma of 60 European patients with HCC or cirrhosis.

Circulating plasma DNA can be retrieved and analyzed for genetic alterations present in the original tumor. In some studies (16–18,20,21), various genetic alterations, such as loss of heterozygosity or K-ras mutations, have been identified in the plasma or sera and tumor tissue from patients with cancer of the lung, head and neck, kidney, colon, and pancreas. These observations and the absence or rarity of alterations in control subjects provide strong evidence that the altered DNA, circulating in the plasma or sera of the patient, originates from the tumor. Although knowledge of the underlying mechanisms of this circulating DNA is still limited, there is some evidence that the DNA is released from the tumor as a

**Table 1.** Characteristics of study participants

Characteristic	Control subjects	Patients with cirrhosis	Patients with hepatocellular carcinoma
From the Gambia			
Total subjects, No.	53	13	53
Median age, y (range)	47 (20–73)	35 (19–68)	46 (19–87)
No. of males (%)	43 (81)	11 (85)	43 (81)
No. of hepatitis B surface antigen-positive subjects (%)	9 (17)	7 (54)	34 (64)
From Europe			
Total subjects, No.		10	50
Median age, y (range)		51 (29–72)	62 (21–85)
No. of males (%)		8 (80)	35 (70)
No. of hepatitis B surface antigen-positive subjects (%)		3 (30)	21 (42)

**Table 2.** Prevalence and adjusted odds ratios (ORs) for Ser-249 p53 mutations in plasma DNA by subject groups from The Gambia\*

	Control subjects†	Patients with cirrhosis	Patients with hepatocellular carcinoma
No. of subjects	53	13	53
No. of Ser-249 p53 mutation-positive subjects (%)	3 (6)	2 (15)	19 (36)
OR (95% confidence interval)†	1.0 (referent)	3.2 (0.4–26.1)	16.4 (3.0–90.5)

\*All sera/plasma samples from European subjects (10 cirrhotic patients and 50 patients with hepatocellular carcinoma) were negative for Ser-249 p53 mutation.

†Control subjects were used as the referent group. Estimated ORs and 95% confidence intervals were adjusted for age, sex, recruitment site, and hepatitis B surface antigen status.



**Table 3.** Comparison of Gambian patients with hepatocellular carcinoma who are negative or positive for the Ser-249 p53 mutation in plasma DNA

	Ser-249 p53 mutation	
	Negative	Positive
No. of subjects	34	19
Median age, y (range)	47.5 (23–87)	46 (19–80)
% male*	31 (91)	12 (63)
No. of hepatitis B surface antigen-positive subjects (%)	23 (68)	11 (58)

\*Two-sided  $P < .05$ . Other two-sided  $P$  values for comparisons between group were not statistically significant. In a model adjusting for age and hepatitis B surface antigen status, the sex difference between mutation-positive and mutation-negative case patients with hepatocellular carcinoma was of borderline statistical significance (two-sided  $P = .053$ ).

glyconucleoprotein complex that may protect it from degradation by nucleases. It remains unclear whether release of tumor DNA into plasma is associated with tumor necrosis, apoptotic cell death, or other selective cellular processes (22–24).

Despite these uncertainties, the data that we present on Ser-249 p53 mutations are consistent with the premise that plasma or serum may be used as a source of tumor-specific DNA. In addition, the data show that this mutation is present in the plasma DNA of patients with HCC from a country with high exposure to AFB<sub>1</sub> but not from Europe. Among 10385 p53 mutations described in human cancers and compiled in the IARC p53 mutation database (<http://www.iarc.fr/p53/>) (25), a total of 197 are Ser-249 p53 mutation, 130 (66%) of which occur in patients with HCC originating from regions with a high incidence of HCC and high exposure to AFB<sub>1</sub>. Twenty-five percent of the Ser-249 p53 mutations were from lung cancers. The presence of this mutation in patients with HCC who are from Europe, the United States, Japan, and Australia is extremely low; only three mutations (one in Europe and two in Japan) were identified among 664 patients with HCC analyzed (6). These data provide strong, albeit indirect, evidence that the DNA containing the Ser-249 p53 mutation originates from the liver. In most developing countries, autopsies or biopsies for HCC are rarely performed; likewise, in this study, we did not have access to tumor tissue from the patients who donated the plasma samples. This is an important limitation of the present study. Collection of paired tumor and plasma samples for confirmation of concordant mutation status is under way in The Gambia. In neighboring Senegal, the prevalence of Ser-249 p53 mutations was reported to be approximately 67% in a series of 15 patients with HCC (26). The

limited data available in The Gambia on the Ser-249 p53 mutation in HCC tumors are consistent with these findings (data not shown). Thus, we estimate that around 70% of patients with HCC who have a detectable Ser-249 p53 mutation in their tumor might also have detectable amounts of mutated DNA in their plasma.

Detection of the Ser-249 p53 mutation in circulating DNA depends on the ability of the method used to detect mutant DNA in a background of wild-type p53 DNA, which is estimated at 1:64 in the present study, based on serial dilution experiments. Preliminary results from the potentially more sensitive mass spectrometry analysis (27,28) of PCR products obtained with different primers also demonstrated the presence of Ser-249 p53 mutation in plasma samples from The Gambia (Friesen M: personal communication). By increasing the sensitivity of this method, DNA with a Ser-249 p53 mutation may be detected in more individuals.

It is interesting that, in this study, Ser-249 p53 mutations were detected not only in patients with HCC but also in patients with cirrhosis (15%) and a small proportion of control subjects (6%). The Ser-249 p53 mutation has been reported previously in nonmalignant liver tissue at frequencies that paralleled the levels of exposure to AFB<sub>1</sub>, suggesting that this mutation could be an early genetic event in hepatocarcinogenesis (29). Detection of the Ser-249 p53 mutation in DNA may thus be useful as a marker of neoplastic development. Alternatively, the presence of the mutation in healthy subjects may reflect chronic exposure to high levels of AFB<sub>1</sub>. The Ser-249 p53 mutation in plasma DNA could be a marker of disease (an early neoplastic effect), exposure (a biologic effect of aflatoxin exposure), or both. Consequently, detection of this mutation may perhaps identify high-risk in-

dividuals who could benefit from more intensive evaluation that could detect HCC earlier.

There is substantial evidence that the Ser-249 p53 mutation present in HCC is directly attributable to DNA damage induced by exposure to dietary AFB<sub>1</sub> (6,30). This proposal is based on the following observations [see (7)]: 1) AFB<sub>1</sub> is highly mutagenic and carcinogenic (3); 2) DNA lesions induced by AFB<sub>1</sub> at guanine residues in double-stranded DNA are by no means random and are dependent on flanking nucleotide sequences (31,32); 3) guanine-to-thymine transversion is the prevalent base change in mutations induced by AFB<sub>1</sub> (33,34); 4) the AFB<sub>1</sub> metabolite binds specifically to the third nucleotide (AGG) in codon 249 (29,35); and 5) the Ser-249 p53 mutation is present in nontumorous tissues in patients from populations at high risk for HCC (29). One recent article (36) that questioned the association between Ser-249 p53 mutations and exposure to AFB<sub>1</sub> was based on a highly selective literature review [see (6,7)] and data from an *in vitro* system that does not mimic the *in vivo* selective advantage of cells containing this specific mutation [see (7)]. At the same time, the Ser-249 p53 mutation has been reported in cultured human cells treated with oxygen radicals (37).

Among patients with HCC, we did not observe any differences in chronic infection with HBV (based on HBsAg status) between those patients with HCC who were positive for the Ser-249 p53 mutation in plasma DNA and patients with HCC who were negative for this mutation (Table 3). Therefore, the highly statistically significant increased risk of HCC associated with Ser-249 p53 mutations in plasma DNA may be, at least in The Gambia, independent of the effects of chronic HBV infection. Geographic and ecologic studies of HCC have been limited by the fact that regions with a high incidence of HCC frequently have a population that is simultaneously exposed to AFB<sub>1</sub> and HBV. Other studies have been limited by the short period of exposure reflected by urinary or serum aflatoxin biomarkers. Because of this complexity, there are at present limited epidemiologic data to permit an estimation of the cancer risk attributable to AFB<sub>1</sub> exposure that is independent of HBV infection and the risk attributable to an interaction between these two factors (38). A recent prospective study of Chi-

nese HBV carriers showed a threefold increased risk for HCC among individuals with detectable aflatoxin levels, as determined by urinary biomarker measurements on pooled urine samples (39,40). This observed effect of aflatoxin with HBV is similar to the independent effect but notably less than the multiplicative interaction associated with aflatoxin biomarkers observed in the original Chinese studies by Ross and colleagues (4,5). Likewise, our data are consistent with an independent causal association between AFB<sub>1</sub> exposure, HBV carrier status, and the subsequent risk of HCC. The use of the Ser-249 p53 mutation detection in plasma or serum DNA as a marker of aflatoxin exposure, along with serologic markers of HBV status, may provide a better estimation of the risk attributable to these agents and a better understanding of their interaction in cancer development. Such studies will have a direct bearing on evaluating the effect of HBV vaccination in preventing HCC presently under way in The Gambia (41).

Despite the 4:1 preponderance of males among patients with HCC, females were consistently more likely to have the Ser-249 p53 mutation. At present, to our knowledge, no data are available to suggest that the prevalence of Ser-249 p53 mutations in HCC tumors might be higher in females than in males. Further studies are needed to confirm this observed sex difference (Table 3).

In addition to p53 mutations, other alterations have been reported in HCC in the genes controlling the transition from G<sub>1</sub> to S phase of the cell cycle, such as amplification of the cyclin D1 gene (42) and inactivation of expression of the p16<sup>INK4</sup> gene, resulting from hypermethylation of this gene (43). A recent study (18) reported a high prevalence of aberrant methylation of the p16 gene in serum DNA from patients with HCC. Somatic mutations in the  $\beta$ -catenin gene (44) and other genetic changes (45) are also frequently detected in HCC tumors. The possibility of detecting these genetic or epigenetic changes in blood specimens from patients with HCC should further elucidate the natural history of HCC and eventually assist in earlier detection and treatment of this cancer.

## REFERENCES

- (1) Parkin DM, Pisani P, Ferlay J. Estimates of the worldwide incidence of 25 major cancers in 1990. *Int J Cancer* 1999;80:827-41.
- (2) International Agency for Research on Cancer (IARC). Hepatitis viruses. Lyon (France): IARC Monogr Eval Carcinog Risks Hum, vol 59; 1994.
- (3) International Agency for Research on Cancer (IARC). Some naturally occurring substances: food items and constituents, heterocyclic aromatic amines and mycotoxins. Lyon (France): IARC Monogr Eval Carcinog Risks Hum, vol 56; 1993. p. 245-395.
- (4) Ross RK, Yuan JM, Yu MC, Wogan GN, Qian GS, Tu JT, et al. Urinary aflatoxin biomarkers and risk of hepatocellular carcinoma. *Lancet* 1992;339:943-6.
- (5) Qian GS, Ross RK, Yu MC, Yuan JM, Gao YT, Henderson BE, et al. A follow-up study of urinary markers of aflatoxin exposure and liver cancer risk in Shanghai, People's Republic of China. *Cancer Epidemiol Biomarkers Prev* 1994;3:3-10.
- (6) Montesano R, Hainaut P, Wild CP. Hepatocellular carcinoma: from gene to public health. *J Natl Cancer Inst* 1997;89:1844-51.
- (7) Hussain SP, Harris CC. Molecular epidemiology of human cancer: contribution of mutation spectra studies of tumor suppressor genes. *Cancer Res* 1998;58:4023-37.
- (8) Hainaut P, Hollstein M. p53 and human cancer: the first ten thousand mutations. *Adv Cancer Res* 2000;77:81-137.
- (9) Hsu IC, Metcalf RA, Sun T, Welsh JA, Wang NJ, Harris CC. Mutational hotspot in the p53 gene in human hepatocellular carcinomas. *Nature* 1991;350:427-8.
- (10) Bressac B, Kew M, Wands J, Ozturk M. Selective G to T mutations of p53 gene in hepatocellular carcinoma from southern Africa. *Nature* 1991;350:429-31.
- (11) Wild CP, Jansen LA, Cova L, Montesano R. Molecular dosimetry of aflatoxin exposure: contribution to understanding the multifactorial etiopathogenesis of primary hepatocellular carcinoma with particular reference to hepatitis B virus. *Environ Health Perspect* 1993;99:115-22.
- (12) Bah E, Hall AJ, Inskip HM. The first 2 years of the Gambian National Cancer Registry. *Br J Cancer* 1990;62:647-50.
- (13) International Agency for Research on Cancer (IARC). The Gambia Hepatitis Intervention Study—annual report, 1997. Lyon (France): IARC; 1998.
- (14) Ryder RW, Whittle HC, Sanneh A, Ajdukiewicz AB, Tulloch S, Yvonne B. Persistent hepatitis B infection and hepatoma in The Gambia, West Africa: a case-control study of 140 adults and their 603 family contacts. *Am J Epidemiol* 1992;136:1122-31.
- (15) Vall Mayans M, Hall AJ, Inskip HA, Chotard J, Lindsay SW, Coromina E, et al. Risk factors for transmission of hepatitis B virus to Gambian children. *Lancet* 1990;336:1107-9.
- (16) Chen XQ, Stroun M, Magnenat JL, Nicod LP, Kurt AM, Lyautey J, et al. Microsatellite alterations in plasma DNA of small cell lung cancer patients. *Nat Med* 1996;2:1033-5.
- (17) Nawroz H, Koch W, Anker P, Stroun M, Sidransky D. Microsatellite alterations in serum DNA of head and neck cancer patients. *Nat Med* 1996;2:1035-7.
- (18) Wong IH, Lo YM, Zhang J, Liew CT, Ng MH, Wong N, et al. Detection of aberrant p16 methylation in the plasma and serum of liver patients. *Cancer Res* 1999;59:71-3.
- (19) Lehman TA, Bennett WP, Metcalf RA, Welsh JA, Ecker J, Modali RV, et al. p53 mutations, ras mutations and p53-heat 70 shock protein complexes in human lung cell lines. *Cancer Res* 1991;51:4090-6.
- (20) Goessl C, Heicappell R, Munker R, Anker P, Stroun M, Krause H, et al. Microsatellite analysis of plasma DNA from patients with clear cell renal carcinoma. *Cancer Res* 1998;58:4728-32.
- (21) Esteller M, Sanchez-Cespedes M, Rosell R, Sidransky D, Baylin SB, Herman JG. Detection of aberrant promoter hypermethylation of tumor suppressor genes in serum DNA from non-small cell lung cancer patients. *Cancer Res* 1999;59:67-70.
- (22) Stroun M, Anker P, Beljanski M, Henri J, Lederrey C, Ojhan M, et al. Presence of RNA in the nucleoprotein complex spontaneously released by human lymphocytes and frog auricles in culture. *Cancer Res* 1978;38:3546-54.
- (23) Anker P, Lefort F, Vasioukhin V, Lyautey J, Lederrey C, Chen XQ, et al. K-ras mutations are found in DNA extracted from the plasma of patients with colorectal cancer. *Gastroenterology* 1997;112:1114-20.
- (24) Stroun M, Anker P, Lyautey J, Lederrey C, Maurice PA. Isolation and characterization of DNA from the plasma of cancer patients. *Eur J Cancer Clin Oncol* 1987;23:707-12.
- (25) Hainaut P, Hernandez T, Robinson A, Rodriguez-Tome P, Flores T, Hollstein M, et al. IARC database of p53 gene mutations in human tumors and cell lines: updated compilation, revised formats and new visualization tools. *Nucleic Acids Res* 1998;26:205-13.
- (26) Coursaget P, Depril N, Chabaud M, Nandi R, Mayelo V, LeCann P, et al. High prevalence of mutations at codon 249 of the p53 gene in hepatocellular carcinomas from Senegal. *Br J Cancer* 1993;67:1395-7.
- (27) Laken SJ, Jackson PE, Kinzler KW, Vogelstein B, Strickland PT, Groopman JD, et al. Genotyping by mass spectrometric analysis of short DNA fragments. *Nat Biotechnol* 1998;13:1352-6.
- (28) Jackson PE, Rashid A, Qian GS, Laken SJ, Friesen MD, Vogelstein B, et al. Specific p53 mutations detected in hepatocellular carcinomas by electrospray mass spectrometry [abstract]. *Proc Am Assoc Cancer Res* 1999;40:619.
- (29) Aguilar F, Harris CC, Sun T, Hollstein M, Cerutti P. Geographic variation of p53 mutational profile in nonmalignant human liver. *Science* 1994;264:1317-9.
- (30) Shimizu Y, Zhu JJ, Han F, Ishikawa T, Oda H. Different frequencies of p53 codon-249 hotspot mutations in hepatocellular carcinomas in Jiang-Su Province of China. *Int J Cancer* 1999;82:187-90.
- (31) Muench KF, Misra RP, Humayun MZ. Sequence specificity in aflatoxin B<sub>1</sub>-DNA interactions. *Proc Natl Acad Sci U S A* 1983;80:6-10.

- (32) Benasutti M, Ejadi S, Whitlow MD, Loechler EL. Mapping the binding site of aflatoxin B1 in DNA: systematic analysis of the reactivity of aflatoxin B1 with guanines in different DNA sequences. *Biochemistry* 1988;27:472-81.
- (33) Foster PL, Eisenstadt E, Miller JH. Base substitution mutations induced by metabolically activated aflatoxin B1. *Proc Natl Acad Sci U S A* 1983;80:2695-8.
- (34) Bailey EA, Iyer RS, Stone MP, Harris TM, Essigmann JM. Mutational properties of the primary aflatoxin B1-DNA adduct. *Proc Natl Acad Sci U S A* 1996;93:1535-9.
- (35) Puisieux A, Lim S, Groopman J, Ozturk M. Selective targeting of p53 gene mutational hotspots in human cancers by etiologically defined carcinogens. *Cancer Res* 1991;51:6185-9.
- (36) Denissenko MF, Koudriakova TB, Smith L, O'Connor TR, Riggs RD, Pfeifer GP. The p53 codon 249 mutational hotspot in hepatocellular carcinoma is not related to selective formation of persistence of aflatoxin B<sub>1</sub> adducts. *Oncogene* 1998;17:3007-14.
- (37) Hussain SP, Aguilar F, Amstad P, Cerutti P. Oxy-radical induced mutagenesis of hotspot codons 248 and 249 of the human p53 gene. *Oncogene* 1994;9:2277-81.
- (38) Wild CP, Hall AJ. Hepatitis B virus and liver cancer: unanswered questions. *Cancer Surveys* 1999;33:35-54.
- (39) Sun Z, Lu P, Gail MH, Pee D, Zhang Q, Ming L, et al. Increased risk of hepatocellular carcinoma in male hepatitis B surface antigen carriers with chronic hepatitis who have detectable urinary aflatoxin metabolite M1. *Hepatology* 1999;30:379-83.
- (40) Wogan GN. Aflatoxin as a human carcinogen. *Hepatology* 1999;30:573-5.
- (41) Viviani S, Jack A, Hall AJ, Maine N, Mendy M, Montesano R, et al. Hepatitis B vaccination in infancy in The Gambia: protection against carriage at 9 years of age. *Vaccine* 1999;17:2946-50.
- (42) Zhang YJ, Jiang W, Chen CJ, Lee CS, Kahn SM, Santella RM, et al. Amplification and overexpression of cyclin D1 in human hepatocellular carcinoma. *Biochem Biophys Res Commun* 1993;196:1010-6.
- (43) Matsuda Y, Ichida T, Matsuzawa J, Sugimura K, Asakura H. p16<sup>INK4</sup> is activated by extensive CpG methylation in human hepatocellular carcinoma. *Gastroenterology* 1999;116:394-400.
- (44) de La Coste A, Romagnolo B, Billuart P, Renard CA, Buendia MA, Soubrane O, et al. Somatic mutations of the beta-catenin gene are frequent in mouse and human hepatocellular carcinomas. *Proc Natl Acad Sci U S A* 1998;95:8847-51.
- (45) Wong N, Lai P, Lee SW, Fan S, Pang E, Liew CT, et al. Assessment of genetic changes in hepatocellular carcinoma by comparative genomic hybridization analysis: relationship to disease stage, tumor size, and cirrhosis. *Am J Pathol* 1999;154:37-43.

## NOTES

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